

HOMOCYSTEINE METABOLISM

J. Selhub

Jean Mayer USDA Human Nutrition Research Center on Aging, Tufts University,
Boston, Massachusetts 02111; e-mail: selhub_vb@hnrc.tufts.edu

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ABSTRACT

Homocysteine is a sulfur amino acid whose metabolism stands at the intersection of two pathways: remethylation to methionine, which requires folate and vitamin B₁₂ (or betaine in an alternative reaction); and transsulfuration to cystathionine, which requires pyridoxal-5'-phosphate. The two pathways are coordinated by *S*-adenosylmethionine, which acts as an allosteric inhibitor of the methylenetetrahydrofolate reductase reaction and as an activator of cystathionine β -synthase. Hyperhomocysteinemia, a condition that recent epidemiological studies have shown to be associated with increased risk of vascular disease, arises from disrupted homocysteine metabolism. Severe hyperhomocysteinemia is due to rare genetic defects resulting in deficiencies in cystathionine beta synthase, methylenetetrahydrofolate reductase, or in enzymes involved in methyl-B₁₂ synthesis and homocysteine methylation. Mild hyperhomocysteinemia seen in fasting conditions is due to mild impairment in the methylation pathway (i.e. folate or B₁₂ deficiencies or methylenetetrahydrofolate reductase thermolability). Post-methionine-load hyperhomocysteinemia may be due to heterozygous cystathionine β -synthase defect or B₆ deficiency. Early studies with nonphysiological high homocysteine levels showed a variety of deleterious effects on endothelial or smooth muscle cells in culture. More recent studies with human beings and animals with mild hyperhomocysteinemia provided encouraging results in the attempt to understand the mechanism that underlies this relationship between mild elevations of plasma homocysteine and vascular disease. The studies with animal models indicated the possibility that the effect of elevated homocysteine is multifactorial, affecting both the vascular wall structure and the blood coagulation system.

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INTRODUCTION

Almost 30 years ago McCully reported that a child suffering from homocystinuria, cystathionuria, and methylmalonic aciduria, secondary to an abnormality of cobalamin metabolism, exhibited arterial lesions that were strikingly similar to those seen in patients with cystathionine β -synthase deficiency (88). This observation led to the proposal that the markedly elevated plasma homocysteine concentrations found in persons with homocystinuria were responsible for the development of premature occlusive vascular disease. In 1976, Wilcken & Wilcken published the first study showing that the homocysteine-cysteine mixed disulfide after a methionine load was significantly higher in patients with coronary artery disease (CAD) than in respective control subjects (150). The importance of their study lies in the fact that the levels of the mixed disulfide seen in these CAD patients, though higher than in healthy subjects, were nevertheless one order of magnitude lower than those seen in homocystinuric patients with congenital defects in cobalamin metabolism or cystathionine β -synthase deficiency. This landmark finding provided the basis for subsequent studies, which since 1990 have increased exponentially (112), and culminated in a meta-analysis published in 1995 (12). This meta-analysis identified a total of 27 studies involving more than 4000 patients with occlusive (cardiovascular, peripheral, and cerebrovascular) vascular disease and the same number of respective control subjects. The data synthesis showed that homocysteine was an independent, graded risk factor for atherosclerotic disease in the coronary, cerebral, and peripheral vessels. A 5- μ M increment in total homocysteine (tHcy) plasma level is associated with an increased risk of 60% for men and 80% for women, of coronary heart disease.

A second review (112) counted a total of 42 additional studies that included ecological cardiovascular disease (CVD) mortality in 11 countries, cross-sectional, case control, nested case control, and cohort populations (112). Only six of these additional studies, including one prospective study, showed no association between homocysteine and disease or mortality. The rest showed positive associations with disease. Another review also suggested that mild hyperhomocysteinemia is associated with an increased risk of thrombotic disease (28). One of the most important studies involved patients with confirmed CAD in which plasma tHcy measured prospectively with a mean follow-up time of 4.6 years (106). After this period, only 3.6% of those with a tHcy level $<9.0 \mu\text{M}$ died, whereas in patients with a tHcy $>15 \mu\text{M}$, the mortality was 24.7%. When a tHcy level below $9 \mu\text{M}$ was used as a reference, mortality rate increased 1.9-, 2.8-, and 4.5-fold among those with tHcy levels of 9–15, 15–20, and $\geq 20 \mu\text{M}$, respectively. A recent study by our group involving the elderly Framingham Study population has demonstrated a twofold increase in all-cause cardiovascular disease mortality among those in the highest quartile of tHcy compared with those in the lowest tHcy quartile (11).

Elevated plasma homocysteine levels were also found to be associated with increased risk of neural tube defects (NTD) (64, 130). A study with a chicken embryo model demonstrated that high homocysteine induces congenital defects of both the neural tube and the heart (117). Other studies demonstrated associations between elevated plasma tHcy and Alzheimer's disease, dementia, and loss of cognitive function (7, 56, 87, 114).

It is important to note that these relations between elevated plasma homocysteine levels and diseases are attained through epidemiological studies. The question of causality remains to be resolved. This chapter reviews homocysteine metabolism with emphasis on its nutritional regulation as it pertains to conditions that are associated with elevated plasma homocysteine levels and discusses current views regarding the mechanism that underlies the relationship between elevated plasma homocysteine levels and vascular disease.

THE HOMOCYSTEINE METABOLIC PATHWAYS

Homocysteine is a non-protein-forming sulfur amino acid whose metabolism is at the intersection of two metabolic pathways: remethylation and transsulfuration (Figure 1). In remethylation, homocysteine acquires a methyl group from N-5-methyltetrahydrofolate or from betaine to form methionine. The reaction with N-5-methyltetrahydrofolate occurs in all tissues and is vitamin B₁₂ dependent, whereas the reaction with betaine is confined mainly to the liver and is vitamin B₁₂ independent. A considerable proportion of methionine is then activated by ATP to form S-adenosylmethionine (SAM). SAM serves primarily as a universal methyl donor to a variety of acceptors. S-adenosylhomocysteine



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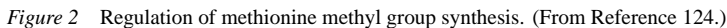
sulfates or is excreted in the urine. Thus, in addition to the synthesis of cysteine, this transsulfuration pathway effectively catabolizes excess homocysteine, which is not required for methyl transfer.

NUTRITIONAL REGULATION OF HOMOCYSTEINE METABOLISM

Studies of the regulation of homocysteine metabolism have demonstrated that the utilization of homocysteine molecules by the transsulfuration and remethylation pathways is nutritionally regulated. Two studies have shown that when the intake of labile methyl groups (i.e. methionine and choline) is modified, the *de novo* synthesis of methionine methyl groups is affected (96, 100). When a basal methionine-containing diet was administered, homocysteine moieties were found to cycle through the remethylation pathway approximately 1.5–2.0 times before being catabolized through the transsulfuration pathway. When dietary methionine was halved, the number of cycles per homocysteine moiety increased twofold. Conversely, when excess dietary methionine was administered, homocysteine cycling fell below basal levels. Similar adaptations to changing levels of dietary methionine were observed in rats (36).

This capacity of the body to discriminate between the remethylation and transsulfuration pathways as a way to adapt to varying amounts of methionine in the diet strongly implies the existence of a coordinate regulation between these two pathways. Available experimental evidence, obtained primarily from measurements of enzyme activities *in vitro*, suggests that this coordination is achieved by at least two mechanisms. The first mechanism is a function of SAM's propensity to act as an allosteric inhibitor of methylenetetrahydrofolate reductase (MTHFR) and as an activator of cystathionine β -synthase (39, 68, 72, 73) (Figures 1 and 2). As such an effector, SAM suppresses the synthesis of an important substrate (N-5-methyltetrahydrofolate) required for remethylation and promotes the initial reaction of transsulfuration (cystathionine synthesis). Thus, intracellular SAM concentration is an important determinant of the fate of homocysteine molecules.

The second mechanism by which remethylation and transsulfuration are coordinated consists of the regulation of intracellular SAM concentration, itself. In the liver, SAM synthesis is catalyzed by two enzymes peculiar to this organ that are immunologically similar but different in other respects (18, 19, 107). One enzyme, a tetramer of high molecular weight, exhibits a high affinity for methionine and is thought to function at normal physiological conditions. The second enzyme is a dimer of a lower molecular weight, has a low affinity for methionine, and is thought to function under conditions of high methionine intake. Thus, changes in intracellular methionine, particularly due to dietary



Additionally, it is also thought that the utilization of SAM is regulated specifically by a reaction in which the methyl group of SAM is transferred to the amino group of glycine, forming sarcosine (Figure 2). This reaction is catalyzed by glycine N-methyltransferase (GNMT), which is abundant in the liver and strongly inhibited by N-5-methyltetrahydrofolate polyglutamates (6, 24, 143). Thus, along with intracellular methionine, N-5-methyltetrahydrofolate participates in the regulation of intracellular SAM concentrations.

1. When dietary methionine is high, the low-molecular-weight SAM synthetase will rapidly convert the incoming methionine to SAM. The resulting rise in intracellular SAM concentration will be associated with (a) inhibition of methylenetetrahydrofolate reductase resulting in suppressed N-5-

methyltetrahydrofolate synthesis, thereby allowing the GNMT enzyme to act near full capacity because of suppressed inhibitor (N-5-methyltetrahydrofolate) concentration; and (b) activation of the cystathionine β -synthase enzyme, thus increasing the rate of homocysteine catabolism. In this way, homocysteine transsulfuration is promoted over remethylation, consistent with the reduced need for de novo methionine synthesis due to the high dietary supply of methionine.

2. Conversely, when the dietary methionine supply is low, SAM concentration is insufficient for the inhibition of MTHFR, resulting in an elevated rate of N-5-methyltetrahydrofolate production. The resulting rise in intracellular N-5-methyltetrahydrofolate concentration will be associated with (a) inhibition of GNMT and thereby conservation of SAM, and (b) an increase in the availability of substrate for homocysteine remethylation. Thus, remethylation will be favored over transsulfuration because the concentration of SAM is too low to activate the cystathionine β -synthase enzyme. This process is consistent with the increased need for de novo methionine synthesis attributed to the low dietary input of methionine.

THE PATHOGENESIS OF HYPERHOMOCYSTEINEMIA

The small amount of homocysteine normally found in the plasma is the result of a cellular export mechanism that complements the catabolism of homocysteine through transsulfuration by helping maintain low intracellular concentrations of this potentially cytotoxic sulfur amino acid (22, 137). Barring kidney malfunction, the occurrence of hyperhomocysteinemia indicates that homocysteine metabolism has in some way been disrupted and that the export mechanism is disposing into the blood excess homocysteine that has accumulated in the cell. This prevents toxicity to the cell but leaves vascular tissue exposed to the possibly deleterious effects of excess homocysteine.

Either a genetic defect in one of the enzymes of homocysteine metabolism or a nutritional deficiency of one or more of the vitamins that participate in homocysteine metabolism can lead to metabolic disruption and potentially to hyperhomocysteinemia. The severity and type of the resulting hyperhomocysteinemia is dependent on the extent to which the particular disturbance affects the coordination of the two pathways of homocysteine metabolism. A discussion of these disturbances follows.

Defective Synthesis of N-5-Methyltetrahydrofolate

Synthesis of N-5-methyltetrahydrofolate is the first step specifically concerned with the synthesis of methionine. An immediate consequence of impaired synthesis of this folate, either because of folate deficiency or because of a

defect in MTHFR, is a depressed synthesis of methionine. This leads to the diversion of homocysteine, which was destined for remethylation, toward the transsulfuration pathway. This latter pathway, however, is incapable of handling the additional homocysteine for two reasons. First, the depressed synthesis of methionine will lead to a decrease in intracellular SAM concentration. Second, the lack of N-5-methyltetrahydrofolate will allow GNMT to be fully active, further decreasing the SAM concentration and increasing the synthesis of homocysteine as a by-product of glycine methylation. Thus, the transsulfuration pathway becomes ineffective because of the increased homocysteine burden in conjunction with a concentration of SAM too low to activate cystathionine synthesis, the initial reaction of transsulfuration. As a result, homocysteine accumulates in the cell and subsequently is exported into the blood, causing hyperhomocysteinemia.

Defective Homocysteine Remethylation

In cases of impaired homocysteine remethylation, as in vitamin B₁₂ deficiency or defects in any of the methyl-cobalamin synthesis enzymes, conditions and consequences are somewhat different from those of impaired N-5-methyltetrahydrofolate synthesis. The methyl trap hypothesis (52) predicts that N-5-methyltetrahydrofolate will actually accumulate when remethylation is impaired. Therefore, despite the decrease in SAM synthesis due to the B₁₂ deficiency or enzyme defect, intracellular SAM concentrations may be less affected because the accumulated N-5-methyltetrahydrofolate will inhibit the utilization of SAM in glycine methylation. As a consequence, less homocysteine will be synthesized from SAM and there will be at least some activation of cystathionine β -synthase. Thus, homocysteinemia that results from impaired homocysteine remethylation may not be as severe as that observed in impaired N-5-methyltetrahydrofolate synthesis because transsulfuration will be somewhat more active in the catabolism of homocysteine. However, it is important to say that hyperhomocysteinemia will occur because homocysteine metabolism is disrupted to a significant extent.

Defective Homocysteine Transsulfuration

No product of the transsulfuration pathway, e.g. cystathionine, cysteine, or taurine, is known to directly affect the remethylation pathway. Nevertheless, the remethylation pathway can be affected by the transsulfuration pathway. When the latter pathway is severely impaired, as in homozygous cystathionine β -synthase defect, there is a diversion of homocysteine toward the remethylation pathway. Under these conditions, the rate of methionine synthesis is increased, leading to a temporal increase in intracellular SAM concentration. This increase

in SAM concentration will continue until the level of this metabolite is sufficient for a feedback inhibition of MTHFR, at which point the remethylation system is inhibited. Consequently, both pathways of homocysteine metabolism are impaired and severe hyperhomocysteinemia results.

When transsulfuration is only mildly impaired, as in vitamin B₆ deficiency or in a heterozygous defect of cystathionine β -synthase, the fully active remethylation pathway and the residual activity of the transsulfuration pathway are sufficient to prevent the precipitation of hyperhomocysteinemia provided the homocysteine burden is low. The homocysteine burden is low when the influx of methionine, the metabolic precursor of homocysteine, is diminished. This occurs under fasting conditions during which there is no dietary input of methionine. However, despite the lack of hyperhomocysteinemia under fasting conditions, there is a disruption of homocysteine metabolism that is detected when the homocysteine burden is increased, i.e. under nonfasting conditions during which there is a significant intake of dietary methionine. This influx of dietary methionine will lead to an increase in intracellular SAM concentration with the following consequences: (a) inhibition of N-5-methyltetrahydrofolate synthesis, hence depressed use of homocysteine through remethylation; and (b) highly active GNMT because of low intracellular N-5-methyltetrahydrofolate concentration, hence accelerated generation of homocysteine through glycine methylation. Coupled with the primary impairment of transsulfuration due to the B₆ deficiency or to the heterozygous cystathionine β -synthase defect, these conditions lead to the significant impairment of the ability of the homocysteine pathways to metabolize homocysteine, and hyperhomocysteinemia results.

Incidentally, this abnormal rise in blood homocysteine concentration after ingestion of a diet containing methionine is the basis for the oral methionine load test. The methionine load test is a comparison of plasma homocysteine concentrations before and after ingestion of a large dose of methionine, usually 100 mg/kg of body weight. The resulting increase in homocysteine synthesis due to the large influx of methionine will test the capacity of the homocysteine pathways. An abnormal test result is attained whenever the capacity of the pathways to metabolize homocysteine is significantly exceeded by the rate of homocysteine generation. In normal individuals, a small rise in blood homocysteine concentration after the load will be observed that returns to baseline within a few hours. In individuals with vitamin B₆ deficiency or a heterozygous cystathionine β -synthase defect, an abnormal response will be observed consisting of a much larger and more persistent rise in blood homocysteine concentration [defined as at least 2 standard deviations (SD) above the rise observed in a normal individual].

SUPPORTING EVIDENCE FOR NUTRITIONAL CAUSES OF HYPERHOMOCYSTEINEMIA

Using rats as models, we found that fasting plasma homocysteine concentrations increased 8- to 10-fold in folate-deficient rats and 2.5-fold in vitamin B₁₂-deficient rats (Figure 3). In the folate-deficient rats, an inverse correlation was observed between intracellular SAM concentration and plasma homocysteine concentration (Figure 4) (91). Moreover, when the folate-deficient rats were administered intraperitoneal (i.p.) injections of ethionine, a methionine analog, plasma homocysteine concentration decreased almost to its normal level (Figure 5). This decrease is thought to be due to *S*-adenosylethionine, which, like SAM, is an effective activator of cystathionine β -synthase, but unlike SAM is less likely to be deethylated and subsequently metabolized to homocysteine (39, 68). This activation by *S*-adenosylethionine reintroduced the coordination

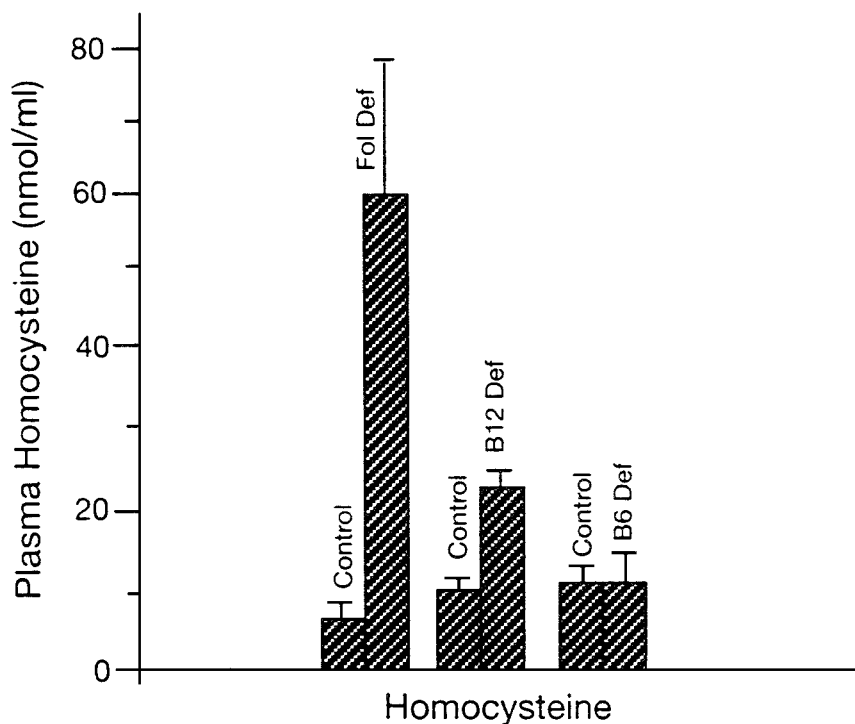


Figure 3 Effects of folate, vitamin B₁₂-, and vitamin B₆-deficiencies on fasting plasma homocysteine levels. (From Reference 124.)

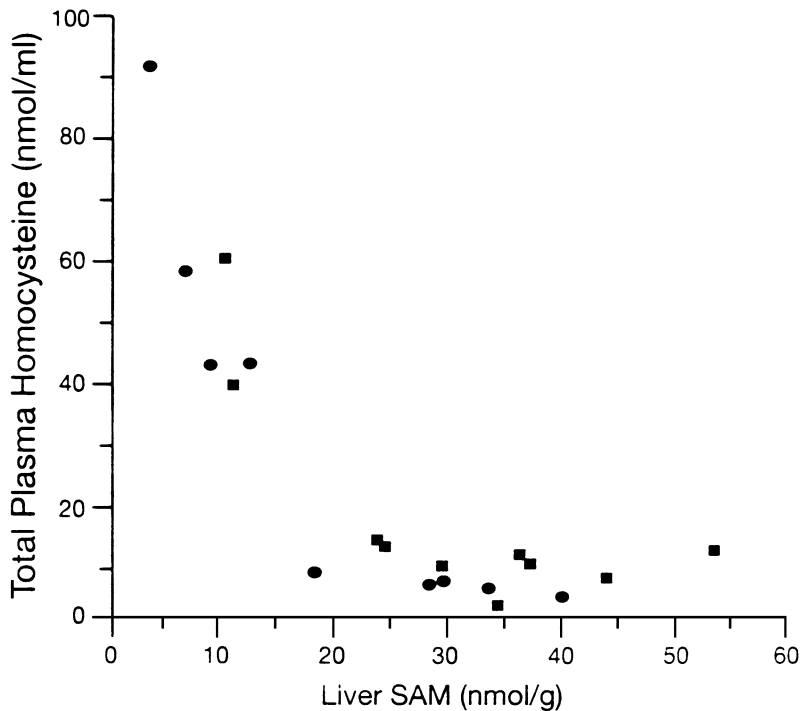


Figure 4 Relationship between fasting plasma homocysteine and hepatic *S*-adenosylmethionine (SAM) concentrations. (From Reference 124.)

between the two pathways that was interrupted in folate deficiency because of diminished SAM synthesis.

In vitamin B₆-deficient rats, fasting plasma homocysteine concentration was not elevated (Figure 3) (92, 127). Moreover, an oral gavage of methionine caused a marked increase in plasma homocysteine concentration in the vitamin B₆-deficient rats that was accompanied by a significant elevation in hepatic SAM concentration (90). This is contrasted with folate-deficient rats in which methionine loading caused no significant change in plasma homocysteine concentration from preload levels (Figure 6) (90).

HYPERHOMOCYSTEINEMIA IN HUMANS

Hyperhomocysteinemia in humans can be distinguished by category, including cause, prevalence, and severity. The more severe cases are due to homozygous

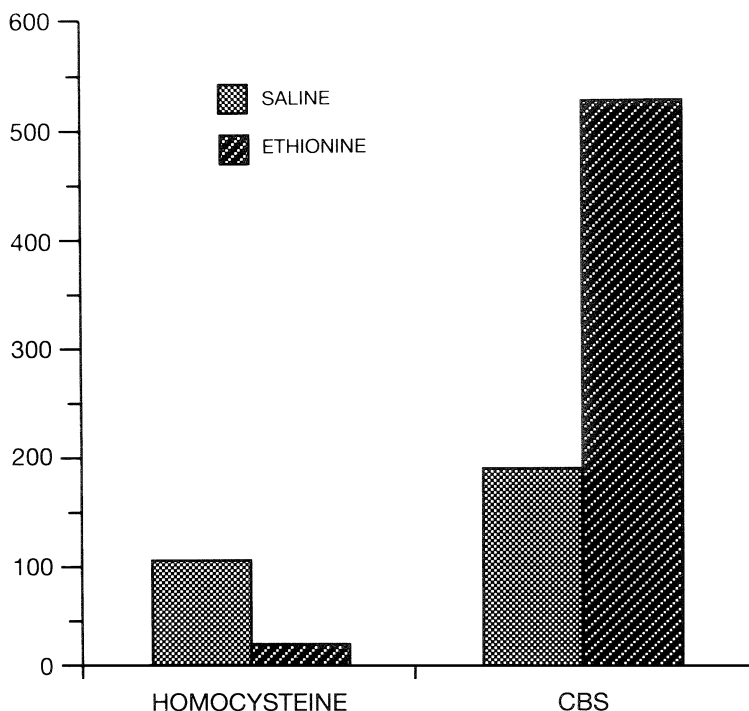


Figure 5 Effect of ethionine administration on fasting plasma homocysteine concentration and hepatic cystathionine β -synthase (C β S) activity in folate-deficient rats. (From Reference 124.)

defects in genes encoding for enzymes of homocysteine metabolism (Table 1). In such cases, a defect in an enzyme involved in either homocysteine remethylation or transsulfuration leads to large elevations of homocysteine in the blood and urine. The classic form of such a disorder—congenital homocystinuria—is caused by homozygous defects in the gene encoding for cystathionine beta synthase (CBS). In these individuals, fasting plasma homocysteine concentrations can be as high as 400 $\mu\text{mol/liter}$ (99). Homozygous defects of other genes that lead to similarly severe elevations in plasma homocysteine include those encoding for MTHFR or for any of the enzymes that participate in the synthesis of methylated vitamin B₁₂ (61, 62, 76, 98, 101, 125).

The more common causes of hyperhomocysteinemia are also moderate in character and may be due to less severe defects in genes encoding for enzymes or from inadequate status of those vitamins that are involved in homocysteine metabolism (Table 1). Plasma homocysteine concentrations in these instances may differ, depending on which arm of the two metabolic pathways

Table 1 Classification of hyperhomocysteinemia^a
Severe hyperhomocysteinemia

High tHcy levels at all times; caused by deficiencies in CBS, MTHFR, or in enzymes of B₁₂ metabolism

Mild hyperhomocysteinemia

Fasting; moderately high tHcy levels under fasting conditions; reflects impaired homocysteine methylation (folate, B₁₂ or moderate enzyme defects, e.g. thermolabile MTHFR)

Post-methionine load

Abnormal increase in tHcy after methionine load. Abnormal net increase reflects impaired homocysteine transsulfuration (heterozygous CBS defects, B₆ deficiency)

^aCystathionine beta synthase (CBS), methylenetetrahydrofolate reductase (MTHFR), total homocysteine (tHcy).

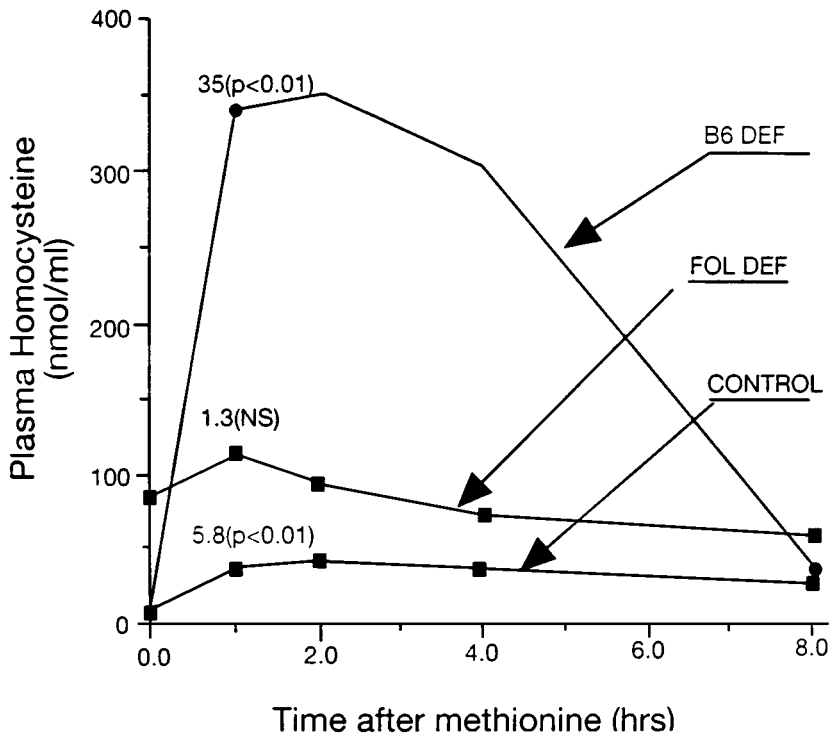


Figure 6 Plasma homocysteine concentrations after methionine loading in control, folate-deficient, and vitamin B₆-deficient rats. (Parentheses) fold increase in homocysteine concentrations from baseline. (From Reference 124.)

of homocysteine metabolism is defective (123). An impairment in the remethylation pathway, even if it is mild, will lead to a substantial increase in plasma homocysteine concentrations under fasting conditions. Such an impairment may be due to inadequate status of either folate or vitamin B₁₂ or to defects in the gene encoding for MTHFR (4, 14, 17, 31, 33, 54, 58, 60, 75, 77, 78, 123, 128, 147, 148). MTHFR contains FAD as a prosthetic group, which raises the possibility that vitamin B₁₂ status is also a determinant of fasting plasma homocysteine levels (126).

In contrast, a mild impairment in the transsulfuration pathway will lead, at most, to a very slight increase in fasting plasma homocysteine levels. This mild impairment, which may be due to heterozygous defects in the CBS gene or inadequate levels of vitamin B₆ (8, 13, 119, 132), is normally identified by an abnormal increase in plasma homocysteine after a methionine loading test or following a meal (90–92, 111, 127).

Evidence of two distinct forms of hyperhomocysteinemia in humans has been derived from preliminary data obtained from 274 consecutive participants in the Family Heart Study (10). Plasma tHcy was measured at fasting and 4 h after a methionine load for each participant. Using homocysteine values greater than 90% for the definition of hyperhomocysteinemia (both fasting and methionine load), it was shown that of 44 hyperhomocysteinemic individuals, 20 (46%) had fasting hyperhomocysteinemia only, 17 (34.5%) had postmethionine load hyperhomocysteinemia only, and just 7 (14%) had both types of hyperhomocysteinemia. Recently, we conducted a placebo-controlled, homocysteine-lowering trial in healthy kidney transplant recipients (9) to show that whereas fasting homocysteine can be lowered by a combination of folate and vitamin B₁₂, post-methionine-load homocysteine can only be lowered by B₆ supplementation.

Existence of an interrelationship between vitamin status and plasma homocysteine was first reported by Kang et al (61), who showed an inverse relationship between homocysteine and plasma folate concentrations. Other studies have shown existence of inverse correlations between homocysteine and folate or vitamin B₁₂ plasma concentrations and the efficacy of vitamin supplementation in the lowering of plasma homocysteine levels (26, 41, 46, 47, 49, 54, 56, 58, 59, 65–72, 104, 110, 135, 140). In these latter studies, folate and vitamin B₁₂, but not vitamin B₆, supplementation reduced fasting plasma homocysteine levels. Vitamin B₆ was found to be effective in lowering post-methionine-load plasma homocysteine.

The independent associations between individual nutrients and homocysteine concentrations were studied in an established cohort of Americans, the Framingham Heart Study (Figure 7) (122). After controlling for age, sex, and other vitamins, nonfasting plasma homocysteine exhibited a strong, nonlinear inverse association with plasma folate. Minimum levels of homocysteine were

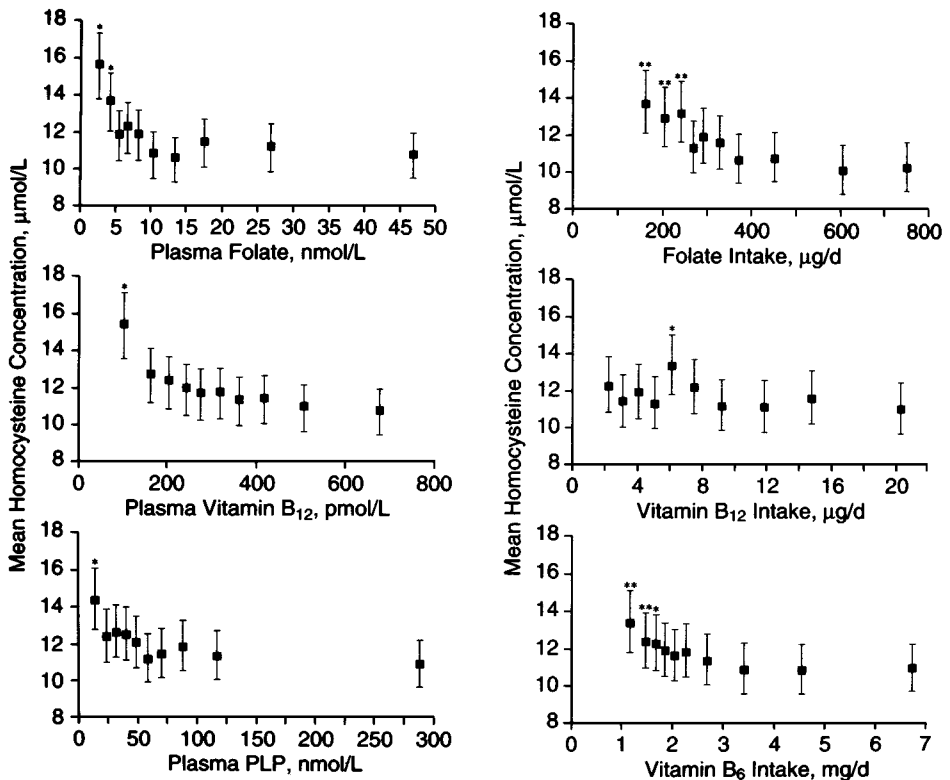


Figure 7 (Left) Mean plasma homocysteine concentrations (and 95% confidence intervals) by deciles of plasma folate (top), vitamin B₁₂ (center), and pyridoxal-5'-phosphate (PLP) (bottom) concentrations. Means are adjusted for age, sex, and other plasma vitamins. (Asterisk) Significant difference from mean in the highest decile ($P < 0.01$). (Right) Mean plasma homocysteine concentrations (and 95% confidence intervals) by deciles of intake of folate (top), vitamin B₆ (center), and vitamin B₁₂ (bottom). Means are adjusted for age, sex, and other vitamin intakes. (Asterisk) indicates significant difference from mean in the highest decile ($P < 0.05$); (double asterisk) significant difference from mean in the highest decile ($P < 0.01$). (From Reference 122.)

observed around 10 nmol/liter of folate and above. Nonfasting plasma homocysteine exhibited weaker inverse associations with plasma concentrations of vitamin B₁₂ and pyridoxal-5'-phosphate. Plasma homocysteine levels also exhibited inverse association with folate intake and, to some extent, vitamin B₆ intake, but not with vitamin B₁₂ intake (Figure 7).

In this elderly Framingham study population (ages 67–96 years), about 30% had elevated plasma homocysteine levels (over 14 μM). Two thirds of the incidences of homocysteine elevations are attributed to inadequate vitamin status or intake.

MTHFR THERMOLABILITY

Relation to Plasma Homocysteine and Folate Status

Mutations that result in severely reduced MTHFR activity and hyperhomocysteinemia are rare (48, 49, 116). However, in 1988, Kang et al (61) reported that two unrelated patients with moderate hyperhomocysteinemia and low folate levels had a variant of MTHFR that was distinguished from the normal enzyme (as measured in lymphocyte extracts) by its lower specific activity (50%) and its thermolability. In subsequent studies, the same group demonstrated that MTHFR thermolability is an inherited recessive trait, which is present in approximately 5% of the general population and 17% of patients with proven coronary artery disease, but it is not associated with neurological complications (57, 59). The cDNA for human MTHFR has recently been isolated (49), and it has been shown that MTHFR thermolability is caused by a point mutation (677C to T transition) at a polymorphic site, resulting in a valine substitution for an alanine in this enzyme (43). The mutation was found in 38% of unselected chromosomes from 57 French Canadian individuals; the homozygous state for the mutation was present in 12% of these subjects and correlated with significantly raised tHcy (43). Preliminary evidence indicates that the frequency of homozygotes for the 677CT mutation may vary significantly in populations from different geographic areas (from 1.4% to 15%) (95).

The impact of MTHFR thermolabile variant on plasma homocysteine levels is unclear. The hyperhomocysteinemia seen in the original patients of Kang et al (61) was associated with low folate plasma levels, and folate supplementation reduced homocysteine to normal levels. In other studies, a large proportion of the population with a TT mutation in the MTHFR gene had normal plasma tHcy levels. Furthermore, the hyperhomocysteinemia seen in those with elevated tHcy was mild and certainly did not correspond with the 50% decrease in enzyme activity seen in vitro with lymphocyte or fibroblast extracts.

This lack of concordance between enzyme activity of the thermolabile MTHFR—determined in cell extracts, and circulating tHcy implied that other factors control the enzyme activity in vivo. In a recent study we demonstrated the occurrence of an interaction between MTHFR thermolability genotype and folate status (55). When plasma folate concentrations were above the median (15.4 nmol/liter), plasma homocysteine levels were low and unrelated to the MTHFR genotype. However, when plasma folate concentrations were below the median, plasma homocysteine levels were significantly higher in homozygotes for the 677CT mutation than in those with the normal genotype (55). The existence of such an interaction has since been confirmed in other studies (2, 21, 121). These data imply that the phenotypic expression of the MTHFR genotypes is dependent on the availability of folate. This suggests

that homozygotes for the thermolabile genotype might have a higher folate requirement than do individuals with a normal genotype.

OTHER FACTORS THAT CONTROL PLASMA HOMOCYSTEINE LEVELS

Plasma homocysteine levels increase with age and are higher in men than in women. High tHcy levels are associated with impaired renal function, high plasma creatinine, smoking, coffee consumption, alcoholism, and certain drugs, including folate antagonists, nitrous oxide, and L-DOPA (112).

PATHOPHYSIOLOGY OF HYPERHOMOCYSTEINEMIA

In spite of the large number of studies indicating that mild elevations of homocysteine in plasma are associated with an increased risk for occlusive vascular disease, thrombosis, and stroke, the question of whether homocysteine per se is responsible for these associations remains unanswered. A survey by Mudd et al (97) of the parents and grandparents of homocystinuric children concluded that heterozygosity for CBS deficiency is not associated with increased risk in heart attacks and stroke (97). Swift & Morrell (131) questioned the validity of some of the methods used by Mudd et al and argued that the data actually point to increased mortality rates in this heterozygote population. Two studies that employed a noninvasive (doppler ultrasound) technique also provided conflicting results. In one, no evidence was found of increased frequency of endothelial plaques in the neck arteries of 25 Irish heterozygotes, compared with 21 control subjects (23). Another study, however, indicated more-frequent early vascular lesions in the iliac and internal carotid arteries in 14 heterozygotes than occurred in 47 controls (118). A study at the molecular biology level (70) examined the CBS alleles in four patients with premature occlusive arterial disease who were (a) hyperhomocysteinemic based on post-methionine-load results and (b) had lower enzyme activity in their fibroblasts (70). None of the eight alleles contained any mutation that resulted in diminished enzyme activity. In a prior study, this same group demonstrated that cultured fibroblasts are not always reliable for testing the phenotypic expression in homocystinuric patients (71). In other studies it was found that prevalence of the more common CBS mutations is not higher in the patient populations (65). In another report, a knockout mouse with CBS deficiency was found to lack manifestations of thrombosis or/and cardiovascular complications and instead exhibited fatty livers (145). This is in spite of the fact that the levels of homocysteine in plasma were 40-fold higher in the homozygote mice than in the control mice. Other inconsistencies relate to the question of whether MTHFR thermolability confers increased risk for

the various diseases. In an earlier study that relied on enzyme activity, a higher prevalence (17%) of this variant was found in a North American CAD population than in healthy control subjects (5%) (57). Similarly, in a Dutch population the prevalence of the 677CT homozygotes was higher in vascular disease patients (15%) than in control subjects (5%); and in an Italian patient population, the prevalence was 29.7% compared with only 15.1% in healthy control subjects (29, 65). A homozygous frequency of 17% in 111 patients with coronary artery disease compared with only 7% in 105 control subjects was reported in Ireland (45). A study conducted in Brazil demonstrated higher prevalence of the 677CT homozygotes in 191 patients with arterial disease (19%) and in 127 patients with venous thrombosis (11%) compared with 296 unmatched control subjects (4%) (5). In Japan, a higher prevalence of the homozygous 677CT mutation was found in 362 patients with coronary artery disease (15.7%) than in 778 controls (10.2%, $P = 0.001$) (94). Furthermore, in patients with triple vessel disease, the prevalence of the 677CT homozygous mutation was 26% compared with only 14% and 15% prevalence in patients with a single or a double vessel disease. Other studies, however, have been contradictory. No difference in the prevalence of the 677CT homozygous mutation was found between coronary artery disease patients and control subjects (see ref 146 for a mini review) in populations from the United States and Australia (120, 149), in a second Dutch population (101), in a French Canadian population (2), in a British population (141), in the Physician Health Study (82), and most recently among women with cardiovascular disease in the United States (121). Of the 17 additional studies that were published in 1998, only four suggested that MTHFR thermolability is a risk factor of vascular disease. The remaining studies found no such association (1, 3, 20, 30, 34, 38, 40, 47, 66, 69, 83, 84, 93, 103, 108, 113, 142).

A recent nested-case-control study of men participating in the Multiple Risk Factor Intervention Trial (MRFIT) found no significant differences in plasma homocysteine levels between control subjects and case patients who had non-fatal myocardial infarctions that occurred within 7 years of sample collection, nor in CHD deaths that occurred more than 11 years after sample collection (40).

Some of these inconsistencies can be explained by differences in genetic background and dietary habits of the study population, or by differences in the pathology among species (e.g. human beings vs mice). Improper selection of the population representing control subjects is likely to account for some of the inconsistencies in the prevalence of the 677CT mutation between patients and control subjects. This may be the case in the original study that reported only a 5% prevalence of the thermolabile MTHFR in the control population (43), and in the first Dutch study, which also reported a 5% prevalence in the control group (29, 65). In other studies from those same countries, prevalence of the

677CT mutation within the control population was reported to be significantly higher. The fact that in the Brazilian study the cases and control population were unmatched (5) casts doubt on the significance of the findings. However, other data that show clear differences in prevalence of the 667CT homozygous mutation between patients and controls (45, 94) cannot be discounted. Rather, these data point to the possibility that the 677CT mutation is indeed a risk factor for vascular disease in certain regions of the world (e.g. Japan, Ireland, and perhaps Brazil). But for the proper interpretation of other inconsistencies, an understanding of the mechanism that underlies the relationship between homocysteine and disease is still required.

It is generally held that different mechanisms are responsible for arterial and venous thromboembolic diseases, and that these mechanisms involve platelet function abnormalities in arterial thrombosis and abnormalities of coagulation and/or fibrinolysis in venous thromboembolism. *Ex vivo* studies looking for such abnormalities in patients with hyperhomocysteinemia have provided inconclusive results (46, 51, 53, 85, 89, 102, 109, 138). In subjects with severe hyperhomocysteinemia due to homozygous CBS deficiency, an abnormally high *in vivo* biosynthesis of thromboxane A₂—as reflected by the urinary excretion of its major metabolite 11-dehydro-thromboxane B₂—has been observed (32).

Administration of aspirin inhibits thromboxane production; urinary appearance returns to baseline levels over a time course consistent with platelet survival, which suggests that platelets are the major source of increased thromboxane urinary excretion. Because thrombin is a potent inducer of platelet activation, the presence of hypercoagulable state was investigated in homocystinuric patients (25). Increased levels of prothrombin fragment 1.2, thrombin-antithrombin complex, and activated protein C were all observed in homocystinuric patients being treated with vitamins who were free of vascular disease. These abnormalities, however, did not correlate with urinary thromboxane excretion. It is interesting to note that protein C levels, but not factor VII and factor II levels, were significantly reduced in homocystinuric patients and correlated with the degree of hyperhomocysteinemia (25). Diet-responsive deficiency of factor VII was previously reported in CBS-deficient patients (15, 85, 109). Reduced protein C levels may at least partly contribute to venous thrombotic manifestations of patients with homozygous CBS deficiency. The observation that the increased urinary thromboxane excretion was independent of homocysteine levels and was present both in vitamin B₆-responsive and -nonresponsive patients may have an impact on treatment of hyperhomocysteinemia (25). It is noteworthy that although the effectiveness of vitamin B₆ in preventing thromboembolism in pyridoxine-responsive patients was shown statistically to be highly significant, the occurrence of thromboembolism was not abolished by vitamin supplementation (37).

Many other studies on the mechanism of hyperhomocysteinemia-related disease employed cell culture systems to show a variety of deleterious effects caused by incubating these cells with homocysteine (28, 86). These effects include the following: inhibition of prostacyclin synthesis, activation of factor V, inhibition of protein C activation, down-regulation of thrombomodulin expression, and blocking of binding of tissue plasminogen activator (t-PA) (but not plasminogen) to endothelial cells. The toxic effect of homocysteine also results in increased platelet adhesion, impaired regulation of endothelium-derived relaxing factor and related nitrogen oxides, induction of tissue factor, suppression of heparan sulfate expression, stimulation of smooth muscle cell proliferation, and oxidation of low-density lipoprotein.

A major shortcoming of these observations is the uncertainty with regard to their pathophysiological significance. Human plasma contains two sulfur-containing amino acids, homocysteine and cysteine, at respective mean normal concentrations of 10 and 240 $\mu\text{mol/liter}$. In mild hyperhomocysteinemia, the levels of homocysteine are on average 30% higher than normal; as opposed to those rare, severe cases of homocystinuria, where the levels may be as high as 200-400 $\mu\text{mol/L}$ (76, 99). Because cysteine appears to be harmless as far as relationship to disease is concerned, these facts strongly imply that the putative action of homocysteine on the blood vessel wall and the coagulation system is stereospecific, involving the entire molecule and taking place at low concentrations (in the micromolar range).

These considerations were not addressed in the various studies on mechanisms. The homocysteine concentrations (1–10 mmol/liter) employed in these studies often exceeded the levels encountered even under the most severe pathological conditions. The possibility that the observed effects seen in these studies were due to nonspecific reactivity of the sulfhydryl group of the homocysteine molecule could not be ruled out. Indeed, in several studies where other thiol-containing compounds such as cysteine and mercaptoethanol were tested, the effects were found to be similar to these seen with homocysteine (for an example see 44; see also 67, 105, 129, 134).

More encouraging results derive from recent studies investigating the impact of moderate hyperhomocysteinemia on a number of hematological parameters in both human beings and laboratory animals. In one important study, high-resolution vascular ultrasonography was used to study endothelium-dependent vasodilation (induced by hyperemia) and endothelium-independent vasodilation (induced by the injection of nitroglycerin) in nonatherosclerotic peripheral conduit arteries of 26 elderly subjects with hyperhomocysteinemia (tHcy = 19.2) and 15 age- and sex-matched normohomocysteinemic control subjects (tHcy = 8.2 μM) (133). The data demonstrated that the endothelium-dependent, but not the endothelium-independent, vasodilation was significantly

lower in the hyperhomocysteinemic group than in the control group. These data were corroborated in a second study that used a cynomolgus monkey model to investigate possible mechanisms of action of mild hyperhomocysteinemia (74). Mild hyperhomocysteinemia was induced by a diet high in methionine, depleted of folate, and free of choline. Total homocysteine concentrations were $10.6 \mu\text{M}$ in the experimental monkeys and $4.0 \mu\text{M}$ in the control group. In response to activation of platelets by infusion of collagen, blood flow to the leg decreased by 42% compared with 14% in the control group. The response of resistance vessels to the endothelium-dependent vasodilators, ADP and acetylcholine, was markedly impaired in the hyperhomocysteinemic monkeys, which indicates that increased vasoconstriction in response to collagen may be caused by decreased vasodilator responsiveness to platelet-generated ADP. Furthermore, thrombomodulin anticoagulant activity in aorta decreased by 34% in the hyperhomocysteinemic monkeys.

The endothelium-dependent vasodilation is a function of the relaxing action of nitric oxide on the blood vessel wall, and demonstration of impaired vasodilation is important as evidence that homocysteine interferes with this action. This possibility was supported by recent studies (63, 80, 139) that used bovine aortic endothelial cells to show that homocysteine treatment is associated with a dose-dependent decrease in NO levels (63, 80, 139).

This decrease in level was not due to decreased activity of endothelial nitric oxide synthase (eNOS) or to decreased activity in *nos3* transcription, which suggests that the decrease in NO levels was not due to decreased synthesis. On the other hand, homocysteine treatment was found to cause a decrease in glutathione peroxidase activity. The authors (63, 80, 139) proposed that homocysteine is a prooxidant that produces H_2O_2 and that under conditions of depressed glutathione peroxidase activity, a greater susceptibility of NO to oxidative inactivation is possible.

Some of these effects of homocysteine on NO and glutathione peroxidase levels in endothelial cell cultures (63) were seen at $10 \mu\text{M}$ DL-homocysteine minimal concentrations and L-cysteine appears not to be effective in these respects. Nevertheless, the idea that homocysteine is functioning as a prooxidant is not in line with its sulfhydryl moiety, which is, rather, considered to be a strong reducing group. A study of the oxidative modification of low-density lipoprotein catalyzed by copper ions, azo compounds, or mediated by mononuclear cells found homocysteine to be ineffective as a prooxidant and suggested that the putative homocysteine-induced atherosclerosis may be explained by mechanisms other than oxidative modification (50).

A study with rats on the effect of folate deficiency on platelet and macrophage activities was done (35). The deficient animals, who were also hyperhomocysteinemic, had peritoneal macrophages with a 20-fold greater tissue factor

activity than the control animals. Folate depletion was also associated with enhanced ADP- and thrombin-induced platelet aggregation. Moreover, the deficient animals exhibited enhanced thromboxane biosynthesis, and there was evidence of diminished unsaturated index in plasma due to a marked fall in long-chain (n-3) polyunsaturated fatty acids. The authors suggested that folic acid deficiency is associated with an increase in oxidative stress, as indicated by an increase in plasma lipid peroxidation and by enhanced susceptibility of erythrocytes to free radicals.

In another recent study, rats were fed a diet containing increasing amounts of homocysteine. The study demonstrated that high plasma homocysteine (8 and 15.2 μM compared with 5.2 μM) is associated with increases in cyclin-dependent kinase in the aortic tissue, lower plasma angiotensin-converting enzyme activity, and higher von Willebrand factor (81).

Rolland et al (115), showed that the hyperhomocysteinemia in minipigs induced by a methionine-rich caseinate diet (9.67 vs 5.64 μM total homocysteine) induced vascular alterations that favor the viscous component of the wall rheology in favor of the elastic component. More important, these alterations share the therapeutic effects by angiotensin-converting enzyme inhibitors in association with hydrochlorothiazides against the atherogenic activation of elastolytic processes.

Although additional studies are required, these emerging data strongly imply that homocysteine may interact with a variety of systems, resulting in different outcomes. Interaction with endothelial cells may result in the impairment of the plasminogenic nature on account of increased thrombogenic properties. On the other hand, interaction with components in the vascular smooth muscle cells may result in enhanced proliferation of these cells and will result in an increased atherogenic tendency (27, 144). However, the possibility that homocysteine is, rather, an indicator of an aberrant metabolism such as localized folate metabolic abnormalities, for example, should still be considered.

TREATMENT OF HYPERHOMOCYSTEINEMIA

Elevations in plasma homocysteine are common in the general population, particularly in the elderly. Vitamin status is a primary determinant of mild-to-moderate hyperhomocysteinemia and accounts for approximately two thirds of all such cases (132). Vitamin supplementation results in near normalization of plasma homocysteine in most cases (16, 42, 79, 136). The meta-analysis of 27 studies by Boushey et al (12) was concerned with the evaluation of the risk of hyperhomocysteinemia for arteriosclerotic vascular disease, with estimation of the reduction of homocysteine levels by folic acid administration, and with the potential reduction of coronary artery disease mortality by increasing folic acid

intake. These analyses proposed that elevations of total homocysteine were an independent graded risk factor for arteriosclerotic vascular disease and calculated that folic acid fortification of food would reduce the annual mortality of the US population by 50,000. Vitamin supplementation may also reduce recurrence of venous thromboembolic disease in patients with hyperhomocysteinemia. The clinical efficacy of this approach has not been tested, however. In addition, the bulk of evidence indicates that fasting total homocysteine determinations may identify up to 50% of the total population of hyperhomocysteinemic subjects. Patients with isolated methionine intolerance may benefit from vitamin B₆ supplementation.

The time is ripe for a placebo/controlled multicenter trial for determining the efficacy of vitamin supplementation in the reduction of morbidity and mortality among patients with occlusive vascular disease, stroke, and thrombosis. Because vitamins are relatively inexpensive, there is little incentive on the part of drug companies to support such a trial, and it is up to government agencies to assume this task. For these reasons it is important that the design of such a trial take into account all the information available. Homocysteine metabolism requires the participation of folate as well as vitamin B₁₂ and vitamin B₆ coenzymes. Reduction of homocysteine levels in plasma requires that all three of these vitamins be supplemented.

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Literature Cited

1. Abbate R, Sardi I, Pepe G, Marcucci R, Brunelli T, et al. 1998. The high prevalence of thermolabile 5–10 methylenetetrahydrofolate reductase (MTHFR) in Italians is not associated to an increased risk for coronary artery disease (CAD). *Thromb. Haemost.* 79:727–30
2. Adams M, Smith PD, Martin D, Thompson JR, Samani NJ. 1996. Genetic analysis of thermolabile methylenetetrahydrofolate reductase as a risk factor for myocardial infarction. *Q. J. Med.* 89:437–44
3. Andrade FL, Annichino-Bizzacchi JM, Saad ST, Costa FF, Arruda VR. 1998. Prothrombin mutant, factor V Leiden, and thermolabile variant of methylenetetrahydrofolate reductase among patients with sickle cell disease in Brazil. *Am. J. Hematol.* 59:46–50
4. Applegarth DA, Hardwick DF, Ingram F, Auckland NL, Bozoian G. 1971. Excretion of S-adenosylmethionine and S-adenosylhomocysteine in homocystinuria. *N. Engl. J. Med.* 285:1265–66
5. Arruda VR, von Zuben PM, Chiaparin LC, Annichino-Bizzacchi JM, Costa FF. 1997. The mutation Ala677→Val in the methylene tetrahydrofolate reductase gene: a risk factor for arterial disease and venous thrombosis. *Thromb. Haemost.* 77:818–21

6. Balaghi M, Horne DW, Wagner C. 1993. Hepatic one-carbon metabolism in early folate deficiency in rats. *Biochem. J.* 291:145–49
7. Bell IR, Edman JS, Selhub J, Morrow FD, Marby DW, et al. 1992. Plasma homocysteine in vascular disease and in nonvascular dementia of depressed elderly people. *Acta Psychiatr. Scand.* 86:386–90
8. Boers GHJ, Fowler B, Smals AGH, Tribbels FJ, Leermakers AI, et al. 1985. Improved identification of heterozygotes for homocystinuria due to cystathionine synthase activity by the combination of methionine loading and enzyme determination in cultured fibroblasts. *Hum. Genet.* 69:164–69
9. Bostom AG, Gohh RY, Beaulieu AJ, Nadeau MR, Hume AL, et al. 1997. Treatment of hyperhomocysteinemia in renal transplant recipients. A randomized, placebo-controlled trial. *Ann. Intern. Med.* 127:1089–92
10. Bostom AG, Jacques PF, Nadeau MR, Williams RR, Ellison RC, Selhub J. 1995. Post-methionine load hyperhomocysteinemia in persons with normal fasting total plasma homocysteine—initial results from the NHLBI Family Heart Study. *Atherosclerosis* 116:147–51
11. Bostom A, Silbershatz H, Jacques P, Selhub J, D'Agostino R, et al. 1999. Non-fasting plasma total homocysteine levels and all-cause cardiovascular disease mortality in elderly Framingham men and women. *Arch. Intern. Med.* In press
12. Boushey CJ, Beresford SA, Omenn GS, Motulsky AG. 1995. A quantitative assessment of plasma homocysteine as a risk factor for vascular disease. Probable benefits of increasing folic acid intakes. *JAMA* 274:1049–57
13. Brattstrom L, Israelsson B, Lindgarde F, Hultberg B. 1988. Higher total plasma homocysteine in vitamin B deficiency than in heterozygosity for homocystinuria due to cystathionine β -synthase deficiency. *Metabolism* 37:175–78
14. Brattstrom L, Israelsson B, Norrving B, Bergqvist D, Throne J, et al. 1990. Impaired homocysteine metabolism in early onset cerebral and peripheral occlusive arterial disease—effects of pyridoxine and folic acid treatment. *Atherosclerosis* 81:51–60
15. Brattstrom L, Israelsson B, Tengborn L, Hultberg B. 1989. Homocysteine, factor VII and antithrombin III in subjects with different gene dosage for cystathionine β -synthase. *J. Inherit. Metab. Dis.* 12:475–82
16. Brattstrom L, Lindgren A, Israelsson B, Andersson A, Hultberg B. 1994. Homocysteine and cysteine: determinants of plasma levels in middle-aged and elderly subjects. *J. Intern. Med.* 236:633–41
17. Brattstrom LE, Israelsson B, Jeppsson J-O, Hultberg BL. 1988. Folic acid—an innocuous means to reduce plasma homocysteine. *Scand. J. Clin. Lab. Invest.* 48:215–21
18. Cabrero C, Martin-Duce A, Ortiz P, Alemany S, Mato JM. 1998. Specific loss of the high molecular weight form of S-adenosyl-L-methionine synthetase in human liver cirrhosis. *Hepatology* 8: 1530–34
19. Cabrero C, Puerta J, Alemany S. 1987. Purification and comparison of two forms of S-adenosyl-L-methionine synthetase from rat liver. *Eur. J. Biochem.* 170:299–304
20. Chapman J, Wang N, Treves TA, Korczyn AD, Bornstein NM. 1998. ACE, MTHFR, factor V Leiden, and APOE polymorphisms in patients with vascular and Alzheimer's dementia. *Stroke* 29:1401–4
21. Christensen B, Frosst P, Lussier-Cacan S, Selhub J, Goyette P, et al. 1997. Correlation of a common mutation in the methylenetetrahydrofolate reductase (MTHFR) gene with plasma homocysteine in patients with premature coronary artery disease. *Arterioscler. Thromb. Vasc. Pathol.* 17:569–73
22. Christensen B, Refsum H, Vintermyr O, Ueland PM. 1991. Homocysteine export from cells cultured in the presence of physiological or superfluous levels of methionine: methionine loading of nontransformed, transformed, proliferating, and quiescent cells in culture. *J. Cell Biol.* 146:52–62
23. Clarke R. 1990. The Irish experience. In *Homocysteinaemia and Vascular Disease*, ed. K Robinson, pp. 41–48. Luxembourg: Comm. Eur. Communities
24. Cook RJ, Wagner C. 1984. Glycine N-methyltransferase is a folate binding protein of rat liver cytosol. *Proc. Natl. Acad. Sci. USA* 81:3631–34
25. Coppola A, Albinini R, Madonna P, Pagano A, Cerbone AM, Di Minno G. 1997. Platelet and monocyte variables in homocystinuria due to cystathionine-beta-synthase deficiency. *Haematologica* 82:189–90
26. Dalery K, Lussier-Cacan S, Selhub J, Davignon J, Latour Y, Genest J. 1995. Homocysteine and coronary artery disease in French Canadian subjects: relationship with vitamin B12, B6, pyri-

- doxal phosphate and folate. *Am. J. Cardiol.* 75:1107–11
27. Dalton M, Gadson P, Wrenn R, Rosenquist TH. 1997. Homocysteine signal cascade. Production of phospholipids, activation of protein C kinase and induction of c-fos and c-myc in smooth muscle cells. *FASEB J.* 11:703–11
 28. D'Angelo A, Selhub J. 1997. Homocysteine and thrombotic disease. *Blood* 90:1–11
 29. de Franchis R, Mancini FP, D'Angelo A, Sebastio G, Fermo I, et al. 1996. Elevated total plasma homocysteine (tHcy) and 677CT mutation of the 5,10-methylenetetrahydrofolate reductase gene in thrombotic vascular disease. *Am. J. Hum. Genet.* 59:262–64
 30. De Stefano V, Chiusolo P, Paciaroni K, Casorelli I, Rossi E, et al. 1998. Prothrombin G20210A mutant genotype is a risk factor for cerebrovascular ischemic disease in young patients. *Blood* 91:3562–65
 31. Dillon MJ, England JM, Gompertz D, Goodey PA, Grant DB, et al. 1974. Mental retardation, megaloblastic anaemia, methylmalonic aciduria and abnormal homocysteine metabolism due to an error in vitamin B12 metabolism. *Clin. Sci. Mol. Med.* 47:43–61
 32. Di Minno G, Davi G, Margaglione M, Cirillo F, Grandone E, et al. 1993. Abnormally high thromboxane biosynthesis in homozygous homocystinuria. Evidence for platelet involvement and Probcuol-sensitive mechanism. *J. Clin. Invest.* 92:1400–6
 33. Dudman NPB, Wilcken DEL, Wang J, Lynch JF, Macey D, Lundberg P. 1993. Disordered methionine/homocysteine metabolism in premature vascular disease: its occurrence, cofactor therapy, and enzymology. *Arterioscler. Thromb.* 13:1253–60
 34. Dunn J, Title LM, Bata I, Johnstone DE, Kirkland SA, et al. 1998. Relation of a common mutation in methylenetetrahydrofolate reductase to plasma homocysteine and early onset coronary artery disease. *Clin. Biochem.* 31:95–100
 35. Durand P, Prost M, Blache D. 1996. Prothrombotic effects of a folic acid deficient diet in rat platelets and macrophages related to elevated homocysteine and decreased n-3 polyunsaturated fatty acids. *Atherosclerosis* 121:231–43
 36. Eloranta TO, Martikainen V, Smith TK. 1990. Adaptation of adenosylmethionine metabolism and methionine recycling to variations in dietary methionine in the rat. *Proc. Soc. Exp. Biol. Med.* 194:364–71
 37. Fenton WA, Rosenberg LE. 1989. Inherited disorders of cobalamin transport and metabolism. In *The Metabolic Basis of Inherited Disease*, ed. CR Scriver, AL Beaudet, WS Sly, D Valle, p. 2065. New York: McGraw-Hill
 38. Fijnheer R, Roest M, Haas FJ, De Groot PG, Derksen RH. 1998. Homocysteine, methylenetetrahydrofolate reductase polymorphism, antiphospholipid antibodies, and thromboembolic events in systemic lupus erythematosus: a retrospective cohort study. *J. Rheumatol.* 25:1737–42
 39. Finkelstein JD, Martin JJ. 1984. Methionine metabolism in mammals: distribution of homocysteine between competing pathways. *J. Biol. Chem.* 259:9508–13
 40. Folsom AR, Nieto FJ, McGovern PG, Tsai MY, Malinow MR, et al. 1998. Prospective study of coronary heart disease incidence in relation to fasting total homocysteine, related genetic polymorphisms, and B vitamins: the Atherosclerosis Risk in Communities (ARIC) study. *Circulation* 98:204–10
 41. Franken DG, Boers GHJ, Blom HJ, Trijbels FJM. 1994. Effect of various regimens of vitamin B-6 and folic acid on mild hyperhomocysteinemia in vascular patients. *J. Inherit. Metab. Dis.* 17:159–62
 42. Franken DG, Boers GHJ, Blom HJ, Trijbels FJM, Kloppenborg PW. 1994. Treatment of mild hyperhomocysteinemia in vascular disease patients. *Arterioscl. Thromb.* 14:465–70
 43. Frosst P, Blom HJ, Milos R, Goyette P, Sheppard CA, et al. 1995. A candidate genetic risk factor for vascular disease: a common mutation in methylenetetrahydrofolate reductase. *Nat. Genet.* 10:111–13
 44. Fryer RH, Wilson BD, Gubler DB, Fitzgerald LA, Rodgers GM. 1993. Homocysteine, a risk factor for premature vascular disease and thrombosis, induces tissue factor activity in endothelial cells. *Arterioscler. Thromb.* 13:1327–33
 45. Gallagher PM, Meleady R, Shields DC, Tan KS, McMaster D, et al. 1996. Homocysteine and risk of premature coronary heart disease. Evidence for a common gene mutation. *Circulation* 94:2154–58
 46. Giannini MJ, Coleman M, Innerfield I. 1975. Antithrombin activity on homocystinuria. *Lancet* 1:1094
 47. Girelli D, Friso S, Trabetti E, Olivieri O, Russo C, et al. 1998. Methylenetetrahy-

- drofolate reductase C677T mutation, plasma homocysteine, and folate in subjects from northern Italy with or without angiographically documented severe coronary atherosclerotic disease: evidence for an important genetic-environmental interaction. *Blood* 91:4158-63
48. Goyette P, Frosst P, Rosenblatt DS, Rozen R. 1995. Seven novel mutations in the methylenetetrahydrofolate reductase gene and genotype/phenotype correlations in severe methylenetetrahydrofolate reductase deficiency. *Am. J. Hum. Genet.* 56:1052-59
 49. Goyette P, Sumner JS, Milos R, Duncan MV, Rosenblatt DS, et al. 1994. Human methylenetetrahydrofolate reductase: isolation of cDNA, mapping and mutation identification. *Nat. Genet.* 7: 195-204
 50. Halvorsen B, Brude I, Drevon CA, Nysom J, Ose L, et al. 1996. Effect of homocysteine on copper ion-catalyzed, azo compound-initiated, and mononuclear cell-mediated oxidative modification of low density lipoprotein. *J. Lipid Res.* 37:1591-600
 51. Harker LA, Slichter SJ, Scott CR, Ross R. 1974. Homocysteinemia. Vascular injury and arterial thrombosis. *N. Engl. J. Med.* 291:537-43
 52. Herbert V, Zalusky R. 1962. Interrelation of vitamin B12 and folic metabolism: folic acid clearance studies. *J. Clin. Invest.* 41:1263-76
 53. Hill-Zobel RL, Pyeritz RE, Scheffel U, Malpica O, Engin S, et al. 1982. Kinetics and distribution of 111-Indium-labeled platelets in patients with homocystinuria. *N. Engl. J. Med.* 307:781-86
 54. Hollowell JG Jr, Hall WK, Coryell ME, McPherson J, Hahn DA. 1969. Homocystinuria and organic aciduria in a patient with vitamin B12 deficiency. *Lancet* 2:1428
 55. Jacques PF, Bostom AG, Williams RR, Ellison RC, Eckfeldt JH, et al. 1996. Relation between folate status, a common mutation in methylenetetrahydrofolate reductase, and plasma homocysteine concentrations. *Circulation* 93:7-9
 56. Joosten E, Lesaffre E, Riezler R, Ghekiere V, Dereymaeker L, et al. 1998. Is metabolic evidence for vitamin B-12 and folate deficiency more frequent in elderly patients with Alzheimer's disease? *J. Gerontol.: Biol. Sci. Med. Sci.* 52:M76-79
 57. Kang SS, Passen EL, Ruggie N, Wong PWK, Sora H. 1993. Thermolabile defect of methylenetetrahydrofolate reductase in coronary artery disease. *Circulation* 88:1463-69
 58. Kang SS, Wong PWK, Norusis M. 1987. Homocysteinemia due to folate deficiency. *Metabolism* 36:458-62
 59. Kang SS, Wong PWK, Susmano A, Sora J, Norusis M, Ruggie N. 1991. Thermolabile methylenetetrahydrofolate reductase: an inherited risk factor for coronary artery disease. *Am. J. Hum. Genet.* 48:536-45
 60. Kang SS, Wong PWK, Zhou JM, Sora J, Lessick M, et al. 1988. Thermolabile methylenetetrahydrofolate reductase in patients with coronary artery disease. *Metabolism* 37:611-13
 61. Kang SS, Zhou J, Wong PWK, Kowalisyn J, Strokosch G. 1988. Intermediate homocysteinemia: a thermolabile variant of methylenetetrahydrofolate reductase. *Am. J. Hum. Genet.* 43:414-21
 62. Kanwar JS, Manaligod JR, Wong PWK. 1976. Morphologic studies in a patient with homocystinuria due to 5,10-methylenetetrahydrofolate reductase deficiency. *Pediatr. Res.* 10:598-609
 63. Keaney JF, Loscalzo J. 1997. Homocyst(e)ine decreases bioavailable nitric oxide by a mechanism involving glutathione peroxidase. *J. Biol. Chem.* 272: 17012-17
 64. Kirke PN, Molloy AM, Daly LE, Burke H, Weir DG, Scott JM. 1993. Maternal plasma folate and vitamin B12 are independent risk factors for neural tube defects. *Q. J. Med.* 86:703-8
 65. Kluijtmans LAJ, van den Heuvel LP, Boers GHJ, Frosst P, Stevens EMB, et al. 1996. Molecular genetic analysis in mild hyperhomocysteinemia: a common mutation in the 5,10-methylenetetrahydrofolate reductase gene is a genetic factor for cardiovascular disease. *Am. J. Hum. Genet.* 58:35-41
 66. Kluijtmans LA, Boers GH, Verbruggen B, Trijbels FJ, Novakova IR, Blom HJ. 1998. Homozygous cystathionine beta-synthase deficiency, combined with factor V Leiden or thermolabile methylenetetrahydrofolate reductase in the risk of venous thrombosis. *Blood* 91:2015-18
 67. Kokame K, Kato H, Miyata T. 1996. Homocysteine-respondent genes in vascular endothelial cells identified by differential display analysis. *J. Biol. Chem.* 271:29659-65
 68. Koracevic D, Djordjevic V. 1977. Effect of trypsin, S-adenosylmethionine and ethionine on L-serine sulphydrase activity. *Experientia* 33:1010-11
 69. Kostulas K, Crisby M, Huang WX, Lannfelt L, Hagenfeldt L, et al. 1998.

- A methylenetetrahydrofolate reductase gene polymorphism in ischaemic stroke and in carotid artery stenosis. *Eur. J. Clin. Invest.* 28:285–89
70. Kozich V, Kraus E, de Franchis R, Fowler B, Boers GHJ, et al. 1995. Hyperhomocysteinemia in premature arterial disease: examination of cystathionine β -synthase alleles at the molecular level. *Hum. Mol. Genet.* 4:623–29
 71. Kozich V, Kraus JP. 1992. Screening for mutations by expressing cDNA segments in *E. Coli*: homocystinuria due to cystathionine β -synthase deficiency. *Hum. Mutat.* 1:113–23
 72. Kutzbach C, Stokstad ELR. 1967. CK inhibition of methylene-tetrahydrofolate reductase in rat liver by S-adenosylmethionine. *Biochim. Biophys. Acta* 139: 217–20
 73. Kutzbach C, Stokstad ELR. 1971. Mammalian methylenetetrahydrofolate reductase: partial purification, properties, and inhibition by S-adenosylmethionine. *Biochim. Biophys. Acta* 250:459–77
 74. Lentz RS, Sobey CG, Pigeors DJ, Bhopatkar MY, Faraci FM, et al. 1996. Vascular dysfunction in monkeys with diet-induced hyperhomocyst(e)inemia. *J. Clin. Invest.* 98:24–29
 75. Levy HL, Cardinale GJ. 1970. Sulfur amino acid abnormalities in experimental vitamin B12 deficiency. *Fed. Proc.* 29:634
 76. Levy HL, Mudd SH, Schulman JD, Dreyfus PM, Abeles RH. 1970. A derangement in B12 metabolism associated with homocysteinemia, cystathioninemia, hypomethioninemia and methylmalonic aciduria. *Am. J. Med.* 48:390–97
 77. Lin JY, Kang SS, Zhou J, Wong PWK. 1989. Homocysteinemia in rats induced by folic acid deficiency. *Life Sci.* 44:319–25
 78. Lindenbaum J, Heaton EB, Savage DG, Brust JC, Garrett TJ, et al. 1988. Neuropsychiatric disorders caused by cobalamin deficiency in the absence of anemia or macrocytosis. *N. Engl. J. Med.* 318:1720–28
 79. Lindgren F, Israelsson B, Lindgren A, Hultberg B, Andersson A, Brattstrom L. 1995. Plasma homocysteine in acute myocardial infarction: homocysteine-lowering effect of folic acid. *J. Intern. Med.* 237:381–88
 80. Loscalzo J. 1996. The oxidant stress of hyperhomocyst(e)inemia. *J. Clin. Invest.* 98:5–7
 81. Lubec B, Labudova O, Hoeger H, Muehl A, Fang-Kircher S, et al. 1996. Homocysteine increases cyclin-dependent kinase in aortic rat tissue. *Circulation* 94:2620–25
 82. Ma J, Stampfer MJ, Hennekens CH, Forst P, Selhub J, et al. 1996. Methylenetetrahydrofolate reductase polymorphism and risk of myocardial infarction in the U.S. physicians. *Circulation* 94:2410–16
 83. Malik NM, Syris P, Schwartzman R, Kaski JC, Crossman DC, et al. 1998. Methylenetetrahydrofolate reductase polymorphism (C-677T) and coronary artery disease. *Clin. Sci.* 95:311–15
 84. Margaglione M, D'Andrea G, d'Addeda M, Giuliani N, Cappucci G, et al. 1998. The methylenetetrahydrofolate reductase TT677 genotype is associated with venous thrombosis independently of the coexistence of the FV Leiden and the prothrombin A20210 mutation. *Thromb. Haemost.* 79:907–11
 85. Maruyama I, Fukuda R, Kaszmama M, Abe T, Yoshida Y, Igata A. 1977. A case of homocystinuria with low antithrombin activity. *Acta Haematol. Jpn.* 40:267–71
 86. Mayer EL, Jacobsen DE, Robinson K. 1996. Homocysteine and coronary atherosclerosis. *J. Am. Coll. Cardiol.* 27:517–27
 87. McCaddon A, Davies G, Hudson P, Tandy S, Cattell H. 1998. Total serum homocysteine in senile dementia of Alzheimer type. *Int. J. Geriatr. Psychiatry* 13:235–39
 88. McCully KS. 1969. Vascular pathology of homocysteinemia: implications for pathogenesis of arteriosclerosis. *Am. J. Pathol.* 56:111–28
 89. Merckx J, Kuntz F. 1981. Deficit en facteur VII et homocystinurie: association fortuite ou syndrome? *Nouv. Presse Med.* 10:3796
 90. Miller JW, Nadeau MR, Smith D, Selhub J. 1994. Vitamin B6 deficiency vs folate deficiency: comparison of responses to methionine loading in rats. *Am. J. Clin. Nutr.* 59:1033–39
 91. Miller JW, Nadeau MR, Smith J, Smith D, Selhub J. 1994. Folate deficiency-induced homocysteinemia in rats: disruption of S-adenosylmethionine's coordinate regulation of homocysteine metabolism. *Biochem. J.* 298:415–19
 92. Miller JW, Ribaya-Mercado JD, Russell RM, Shepard DC, Morrow FD, et al. 1992. Total homocysteine in fasting plasma is not a good indicator of B6 deficiency. *Am. J. Clin. Nutr.* 55:1154–60
 93. Morita H, Kurihara H, Tsubaki S, Sugiyama T, Hamada C, et al. 1998. Methylenetetrahydrofolate reductase gene polymorphism and ischemic stroke in

- Japanese. *Arterioscler. Thromb. Vasc. Biol.* 18:1465–69
94. Morita H, Taguchi J, Kurihara H, Kitaoka M, Kaneda H, et al. 1997. Genetic polymorphism of 5,10-methylenetetrahydrofolate reductase (MTHFR) as a risk factor for coronary artery disease. *Circulation* 95:2032–36
 95. Motulsky A. 1996. Nutritional ecogenetics: homocysteine-related arteriosclerotic vascular disease, neural tube defects, and folic acid. *Am. J. Hum. Genet.* 58:17–20
 96. Mudd SH, Ebert MH, Scriver CR. 1980. Labile methyl group balances in the human: the role of sarcosine. *Metabolism* 29:707–20
 97. Mudd SH, Havlik R, Levy HL, McKusick VA, Feinleib M. 1981. A study of cardiovascular risk in heterozygotes for homocystinuria. *Am. J. Hum. Genet.* 33:883–93
 98. Mudd SH, Levy HL, Morrow G. 1970. Deranged B12 metabolism: effects on sulfur amino acid metabolism. *Biochem. Med.* 4:193–214
 99. Mudd SH, Levy HL, Skovby F. 1995. Disorders of transsulfuration. In *The Metabolic and Molecular Basis of Inherited Disease*, ed. CR Scriver, AL Beaudet, WS Sly, D Valle, pp. 1279–327. New York: McGraw-Hill
 100. Mudd SH, Poole JR. 1975. Labile methyl balances for normal humans on various dietary regimens. *Metabolism* 24:721–35
 101. Mudd SH, Uhlenendorf BW, Freeman JM, Finkelstein JD, Shih VE. 1972. Homocystinuria associated with decreased methylenetetrahydrofolate reductase activity. *Biochem. Biophys. Res. Commun.* 46:905–12
 102. Munnich A, Saudbray JM, Dautzenberg MD, Parvy P, Ogier H, et al. 1983. Diet-responsive proconvertin (factor VII) deficiency in homocystinuria. *J. Pediatr.* 102:730–34
 103. Nakata Y, Katsuya T, Takami S, Sato N, Fu Y, et al. 1998. Methylenetetrahydrofolate reductase gene polymorphism: relation to blood pressure and cerebrovascular disease. *Am. J. Hypertens.* 11:1019–23
 104. Naurath HJ, Joosten E, Riezler R, Stabler SP, Allen RH, Lindenbaum J. 1995. Effects of vitamin B12, folate, and vitamin B6 supplements in elderly people with normal serum vitamin concentrations. *Lancet* 346:858–59
 105. Nishinaga M, Ozawa T, Shimada K. 1993. Homocysteine, a thrombogenic agent, suppresses anticoagulant heparan sulfate expression in cultured porcine aortic endothelial cells. *J. Clin. Invest.* 92:1381–86
 106. Nygard O, Nordrehaug JE, Refsum H, Ueland PM, Farstad M, Vollset SE. 1997. Plasma homocysteine levels and mortality in patients with coronary artery disease. *N. Engl. J. Med.* 337:230–36
 107. Okada G, Teraoka H, Tsukada K. 1981. Multiple species of mammalian S-adenosylmethionine synthase. Partial purification and characterization. *Biochemistry* 20:934–40
 108. Ou T, Yamakawa-Kobayashi K, Arinami T, Amemiya H, Fujiwara H, et al. 1998. Methylenetetrahydrofolate reductase and apolipoprotein E polymorphisms are independent risk factors for coronary heart disease in Japanese: a case-control study. *Atherosclerosis* 137:23–28
 109. Palareti G, Salardi S, Pizzi S, Legnani C, Poggi M, et al. 1986. Blood coagulation changes in homocystinuria: effects of pyridoxine and other specific therapy. *J. Pediatr.* 109:1001–6
 110. Pancharunty N, Lewis CA, Sauberlich HE, Perkins LL, Go RCO, et al. 1994. Plasma homocysteine, folate and B6 concentrations and the risk for early-onset coronary artery disease. *Am. J. Clin. Nutr.* 59:940–48
 111. Park YK, Linkswiler H. 1970. Effect of vitamin B6 depletion in adult man on the excretion of cystathionine and other methionine metabolites. *J. Nutr.* 100:110–16
 112. Refsum H, Ueland PM, Nygard O, Vollset SE. 1998. Homocysteine and cardiovascular disease. *Annu. Rev. Med.* 49:31–62
 113. Reinhardt D, Sigusch HH, Vogt SF, Farker K, Muller S, Hoffmann A. 1998. Absence of association between a common mutation in the methylenetetrahydrofolate reductase gene and the risk of coronary artery disease. *Eur. J. Clin. Invest.* 28:20–23
 114. Riggs KM, Spiro A, Tucker K, Rush D. 1996. Relations of vitamin B-12, vitamin B-6, folate, and homocysteine to cognitive performance in the normative aging study. *Am. J. Clin. Nutr.* 63:306–14
 115. Rolland PH, Friggi A, Barlatier A, Piquet P, Latrille V, et al. 1995. Hyperhomocysteinemia induced vascular damage in minipig. Captopril-hydrochlorotiazide combination prevents elastic alterations. *Circulation* 91:1161–74
 116. Rosenblatt DS. 1989. Inherited disorders of folate transport and metabolism. See Ref. 37, pp. 2049–64
 117. Rosenquist TH, Ratashak SA, Selhub J. 1996. Homocysteine induces congenital defects of the heart and neural tube: effect of folic acid. *Proc. Natl. Acad. Sci. USA* 93:15227–32

118. Rubba P, Faccenda F, Pauciuolo P, Carbone L, Mancini M, et al. 1990. Early signs of vascular disease in homocystinuria: a non-invasive study by ultrasound methods in eight families with cystathionine- β -synthase deficiency. *Metabolism* 39:1191-95
119. Sardharwalla IB, Fowler B, Robins AJ, Komrower GM. 1974. Detection of heterozygotes for homocystinuria. *Arch. Dis. Child.* 49:553-59
120. Schmitz C, Lindpaintner K, Verhoef P, Gaziano JM, Buring J. 1996. Genetic polymorphism of methylenetetrahydrofolate reductase and myocardial infarction: a case-control study. *Circulation* 94:1812-14
121. Schwartz SM, Siscovick DS, Malinow MR, Rosendaal FR, Beverly RK, et al. 1997. Myocardial infarction in young women in relation to plasma total homocysteine, folate, and a common variant in the methylenetetrahydrofolate reductase gene. *Circulation* 96:412-17
122. Selhub J, Jacques PF, Wilson PWF, Rush D, Rosenberg IH. 1993. Vitamin status and intake as primary determinants of homocysteinemia in the elderly. *JAMA* 270:2693-98
123. Selhub J, Miller JW. 1991. The pathogenesis of homocysteinemia: interruption of the coordinate regulation by S-adenosylmethionine of the remethylation and transsulfuration of homocysteine. *Am. J. Clin. Nutr.* 55:131-38
124. Selhub J, Miller JW. 1994. Regulation of plasma homocysteine concentration by nutrients and drugs. In *Methionine Metabolism: Molecular Mechanisms and Clinical Implications*, ed. JM Mato, A Caballero, pp. 89-98. Madrid: Cons. Super. de Invest. Cient.
125. Shih VE, Salam MZ, Mudd SH, Uhlenhuth BW, Adams RD. 1972. A new form of homocystinuria due to 5,10-methylenetetrahydrofolate reductase deficiency. *Pediatr. Res.* 6:395
126. Shimakawa T, Nieto FJ, Malinow MR, Chambless LE, Schreiner PJ, Szklo M. 1997. Vitamin intake: a possible determinant of plasma homocyst(e)ine among middle-aged adults. *Ann. Epidemiol.* 7: 285-93
127. Smolin LA, Benevenga NJ. 1984. Factors affecting the accumulation of homocyst(e)ine in rats deficient in vitamin B6. *J. Nutr.* 114:103-11
128. Stabler SP, Marcell PD, Podell ER, Allen RH, Savage DG, Lindenbaum J. 1988. Elevation of total homocysteine in the serum of patients with cobalamin or folate deficiency detected by capillary gas-chromatography-mass spectrometry. *J. Clin. Invest.* 81:466-74
129. Stamler JS, Osborne JA, Jaraki M, Rabini LE, Mullins M, et al. 1993. Adverse vascular effects of homocysteine are modulated by endothelium-derived relaxing factor and related oxides of nitrogen. *J. Clin. Invest.* 91:308-18
130. Steegers-Theunissen RP, Boers GH, Blom HJ, Nijhuis JG, Thomas CMG, et al. 1995. Neural tube defects and elevated homocysteine levels in amniotic fluid. *Am. J. Obstet. Gynecol.* 172:1436-41
131. Swift M, Morrell D. 1982. Cardiovascular risk in homocystinuria family members. *Am. J. Hum. Genet.* 34:1016-18
132. Swift ME, Schultz TD. 1986. Relationship of vitamins B6 and B12 to homocysteine levels: risk for coronary heart disease. *Nutr. Rep. Int.* 34:1-14
133. Tawakol A, Omland T, Gerhard M, Wu JT, Creager MA. 1997. Hyperhomocyst(e)inemia is associated with impaired endothelium-dependent vasodilation in humans. *Circulation* 95:1119-21
134. Tsai J-C, Perrella MA, Yoshizumi M, Hsieh CM, Haber E, et al. 1994. Promotion of vascular smooth muscle cell growth by homocysteine: a link to atherosclerosis. *Proc. Natl. Acad. Sci. USA* 91:6369-73
135. Ubbink JB, Vermaak WJH, van der Merwe A, Becker PJ. 1993. Vitamin B12, vitamin B6, and folate nutritional status in men with hyperhomocysteinemia. *Am. J. Clin. Nutr.* 57:47-53
136. Ubbink JB, Vermaak WJH, van der Merwe A, Becker PJ, Delpont R, Potgieter HC. 1994. Vitamin requirements for the treatment of hyperhomocysteinemia in humans. *J. Nutr.* 124:1927-33
137. Ueland PM, Refsum H, Male R, Lillehaug JR. 1986. Disposition of endogenous homocysteine by mouse fibroblast C3H/10T1/2 Cl 8 and the chemically transformed C3H/10T1/2 MCA Cl 16 cells following methotrexate exposure. *J. Natl. Cancer Inst.* 77:283-89
138. Uhlemann ER, TenPas JH, Lucky AW, Schulman JD, Mudd SH, Shulman NR. 1976. Platelet survival and morphology in homocystinuria due to cystathionine synthase deficiency. *N. Engl. J. Med.* 295: 1283-86
139. Upchurch GR Jr, Welch GN, Freedman JE, Loscalzo J. 1995. Homocysteine attenuates endothelial glutathione peroxidase and thereby potentiates peroxide-mediated cell injury. *Circulation* 92:1086

140. van den Berg N, Franken DG, Boers GHJ, Blom HJ, Jakobs C, et al. 1994. Combined vitamin B-6 plus folic acid therapy in young patients with arteriosclerosis and hyperhomocysteinemia. *J. Vasc. Surg.* 20:933–40
141. Verhoef P, Kok FJ, Kluijtmans LA, Blom HJ, Refsum H, et al. 1997. The 677C-T mutation in the methylenetetrahydrofolate reductase gene: associations with plasma total homocysteine levels and risk of coronary atherosclerotic disease. *Atherosclerosis* 13:105–13
142. Verhoef P, Rimm EB, Hunter DJ, Chen J, Willett WC, et al. 1998. A common mutation in the methylenetetrahydrofolate reductase gene and risk of coronary heart disease: results among U.S. men. *J. Am. Coll. Cardiol.* 32:353–59
143. Wagner C, Briggs WT, Cook RJ. 1985. Inhibition of glycine N-methyltransferase activity by folate derivatives: implications for regulation of methyl group metabolism. *Biochem. Biophys. Res. Commun.* 127:746–52
144. Wang H, Yoshizumi M, Lai KH, Tsai JC, Perrella MA, et al. 1997. Inhibition of growth and p21ras methylation in vascular endothelial cells by homocysteine but not cysteine. *J. Biol. Chem.* 272:25380–85
145. Watanabe M, Osada J, Aratani Y, Kluckman K, Reddick R, et al. 1995. Mice deficient in cystathionine β -synthase: animal models for mild and severe homocyst(e)inemia. *Proc. Natl. Acad. Sci. USA* 92:1585–89
146. Wilcken DEL. 1997. MTHFR 677C-T mutation, folate intake, neural-tube defect, and risk of cardiovascular disease. *Lancet* 350:603–4
147. Wilcken DEL, Dudman NPB, Tyrrell PA, Robertson MR. 1988. Folic acid lowers elevated plasma homocysteine in chronic renal insufficiency: possible implications for prevention of vascular disease. *Metabolism* 37:697–701
148. Wilcken DEL, Gupta VJ, Betts AK. 1981. Homocysteine in the plasma of renal transplant recipients: effects of cofactors for methionine metabolism. *Clin. Sci.* 61:743–49
149. Wilcken DEL, Wang XL, Sim AS, McCredie RM. 1996. Distribution in healthy and coronary populations of the methylenetetrahydrofolate reductase (MTHFR) C677T mutation. *Arterioscler. Thromb. Vasc. Biol.* 16:878–82
150. Wilcken DEL, Wilcken B. 1976. The pathogenesis of coronary artery disease. A possible role for methionine metabolism. *J. Clin. Invest.* 57:1079–82